

Review **Multifunctional Role of** *Acca sellowiana* **from Farm Management to Postharvest Life: A Review**

Thomas Vatrano ¹ , Margherita Amenta ² [,](https://orcid.org/0000-0002-4415-8470) Andrea Copetta ³ [,](https://orcid.org/0000-0001-6219-7206) Maria Guardo ² , Angelina Nunziata ⁴ [,](https://orcid.org/0000-0002-9946-7334) Maria Concetta Strano [2](https://orcid.org/0000-0001-7152-5064) and Milena Petriccione 4,*

- 1 Independent Researcher, 88021 Borgia, Italy; thomasvatrano@gmail.com
- ² CREA-Research Centre for Olive, Fruit and Citrus Crops, 95024 Acireale, Italy; margherita.amenta@crea.gov.it (M.A.); maria.guardo@crea.gov.it (M.G.); mariaconcetta.strano@crea.gov.it (M.C.S.)
- ³ CREA-Research Centre Vegetable and Ornamental Crops, 18038 Sanremo, Italy; andrea.copetta@crea.gov.it
- ⁴ CREA-Research Centre for Olive, Fruit and Citrus Crops, 81100 Caserta, Italy; angelina.nunziata@crea.gov.it
- ***** Correspondence: milena.petriccione@crea.gov.it; Tel.: +39-082-325-6244

Abstract: Feijoa (*Acca sellowiana* (O. Berg) Burret syn *Feijoa sellowiana* Berg) is a monotypic species belonging to the Myrtaceae family. Feijoa is cultivated in different countries, and it is mainly consumed as fresh fruit, due to its attractive nutritional quality, nevertheless, several feijoa-based products have been produced and are available in some niche and local markets. The fruits are not the only edible part of the tree since petals of hermaphrodite showy flowers can also be eaten. Fruits are rich in vitamin C, fiber, potassium, iodine, phosphorus, sugars, and calcium. In addition, a large number of bioactive compounds, with many health-promoting benefits, have been identified. This review is carried out with the aim to provide comprehensive and updated over-view of the state of the art related to the knowledge on feijoa, evaluating the main agronomic, qualitative, and nutraceutical traits of its edible parts. Moreover, the variability in feijoa genetic resources to identify and select promising genotypes useful for breeding programs and the postharvest management of fruit and edible flowers will be discussed. Overall, feijoa has great potential to be considered as a sustainable fruit crop from farm to table.

Keywords: feijoa; edible flowers; agronomy; nutraceutical; genetic; postharvest

1. Introduction

Feijoa tree (*Acca sellowiana* (O. Berg) Burret syn *Feijoa sellowiana* Berg.) native of extreme southern Brazil, northern Argentina, western Paraguay, and Uruguay, is now widely cultivated in different countries of the world such as USA, France, Italy, Turkey, Iran, Australia, and New Zealand for the production of its fruit or as ornamental tree [\[1](#page-9-0)[,2\]](#page-9-1). Feijoa remains a relatively underutilized species compared to other fruit crops, but, thanks to its easy adaptability in subtropical regions, it could be cultivated in other countries and commercialized as a new high-value fruit crop for its fruit and edible flowers [\[3\]](#page-9-2). This crop also has good development prospects in organic farming based on its high adaptability to different agronomic conditions and low susceptibility to pests and diseases [\[4\]](#page-9-3).

Feijoa is an exotic species with a high economical potential for its functional fruits with sweet and aromatic flavor and a high level of bioactive compounds which promote numer-ous benefits to human health and/or reduce the risk of developing chronic diseases [\[3](#page-9-2)[,5](#page-9-4)[,6\]](#page-9-5). Furthermore, the sweet taste of the feijoa petals makes them interesting for the supply chain of edible flowers [\[7\]](#page-9-6).

Feijoa peel represents about 60% (*w*/*w*) of total fruit weight and it is a rich source of functional bioactive compounds that is discarded during the industrial processing or consumption of whole fresh fruits [\[8,](#page-9-7)[9\]](#page-9-8). This by-product still has relevant chemical and nutritional value and could be upcycled for human consumption, being a potential

Citation: Vatrano, T.; Amenta, M.; Copetta, A.; Guardo, M.; Nunziata, A.; Strano, M.C.; Petriccione, M. Multifunctional Role of *Acca sellowiana* from Farm Management to Postharvest Life: A Review. *Agronomy* **2022**, *12*, 1802. [https://](https://doi.org/10.3390/agronomy12081802) doi.org/10.3390/agronomy12081802

Academic Editor: Anastasios Darras

Received: 3 July 2022 Accepted: 27 July 2022 Published: 29 July 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/) $4.0/$).

source of functional ingredients for industrial applications in the food and pharmaceutical industries [\[10\]](#page-9-9).

that plants take up for an annual yield of 20 t ha**−1**, which is: 26 kg (N); 1.5 kg (P); 41 kg

This review was created with the aim to provide a comprehensive and updated overview of the recent advances on feijoa from agronomic features to nutraceutical traits of feijoa flowers and fruits. The postharvest management and the reuse of processing waste are also reviewed. **data spots of interviewed** and can cause enormous losses in contract α

2. Biological and Agronomic Features stem rot and the root rot due to fungi of genus *Phytophthora*, *Pythium,* and *Rosellinia*.

Feijoa is a slow-growing evergreen tree that naturally grows to a height of 4–6 m [\[4\]](#page-9-3) (Figure [1A](#page-1-0)). The branches have opposite persistent, elliptical, and leathery leaves, bright green on the upper side and whitish on the underside [11]. The showy and hermaphrodite flowers, inserted on the current season's shoot, are single or in a cluster [\[12](#page-9-11)[,13\]](#page-9-12).

Figure 1. Feijoa experimental orchard located in Caserta, Southern Italy (41°90′ N, 14°80′ E) and owned by the CREA-OFA (Italy) (**A**), flower (**B**), and fruits cv Mammouth (**C**). owned by the CREA-OFA (Italy) (**A**), flower (**B**), and fruits cv Mammouth (**C**).

Flowering period occurs in spring, but it is variable depending on the cultivars and environmental conditions [\[13\]](#page-9-12) (Figure [1B](#page-1-0)). In the Mediterranean area, floral development
and emergence last for two menths (Mid-Anril to June) and this was also senfirmed in where the soft juice soft into $\frac{1}{2}$ into the small seeds and contained $\frac{1}{2}$ into the groups based on $\frac{1}{2}$ into the groups based on $\frac{1}{2}$ is a contained by $\frac{1}{2}$ in $\frac{1}{2}$ in $\frac{1}{2}$ in $\frac{1}{2}$ Chinese feijoa orchards by Zou et al. [\[14\]](#page-9-13). Feijoa cultivars bear different barriers to selfand emergence last for two months (Mid-April to June) and this was also confirmed in

fertilization, some of them are strictly self-incompatible, while others are compatible or partially compatible [\[13\]](#page-9-12). Cross-pollination increases a fruit set up to 80–90%, while selfpollination halves it; this is due to a higher percentage of abscission in self-pollinated than in cross-pollinated flowers [\[15\]](#page-9-14).

Fruit shows persistent calyx with four sepals with variable shape and weight $(40-150 \text{ g})$ [\[4\]](#page-9-3) (Figure [1C](#page-1-0)). Skin berry may be smooth or rough and wraps a white granular flesh made up of sclereids in the outer parenchymatous region that becomes brown in color at ripeness [\[16\]](#page-9-15). Small and flattened seeds are immersed in a jelly substance placed in a central cavity constituted of about four locules [\[4\]](#page-9-3). Fruit ripening occurs from March to June in the southern hemisphere and from October to December in the northern hemisphere [\[4,](#page-9-3)[6\]](#page-9-5). The fruits have an intense and aromatic flavor reminiscent of strawberry and pineapple and are mainly consumed as a fresh product.

Nowadays, feijoa is cultivated in different countries and the optimization of crop management conditions has allowed to improve its bio-agronomic performances obtaining productions that reach 20 t ha⁻¹ or more in commercial orchards [\[17\]](#page-9-16). For optimal crop management conditions, it is recommended to always use different feijoa cultivars to overcome self-sterility barriers. The planting densities are influenced by cultivar vigor ranging from 400 trees ha⁻¹ (more vigorous) to almost 880 trees ha⁻¹ (less vigorous) with 5 m and 4.5 m between rows and 5 and 2.5 m between trees, respectively. Feijoa needs a well-drained and slightly acidic (pH 6.0–6.5) soil with a high organic matter content [\[18](#page-9-17)[,19\]](#page-9-18). This species grows well in free exposure to direct solar radiation of \geq 1500 h (direct sunlight per year) up to 700–1000 m.a.s.l., but in Colombia, it is well-adapted until 1800–2700 m.a.s.l. [\[17\]](#page-9-16). Quintero-Castillo [\[20\]](#page-9-19) demonstrated that good production can be obtained in areas where there are 1000 h/year of direct sunlight with favorable pedoclimatic conditions. Furthermore, the production of saplings in nurseries and orchards of this species should be under full sun or up to 30% shade, because it makes saplings more vigorous and can anticipate and improve fruit production [\[21\]](#page-9-20).

Feijoa tolerates winter frosts, and temperatures down to −4 ◦C do not cause significant damage in adult plants [\[11\]](#page-9-10); it requires a certain number of chilling hours below 7 °C to produce a major number of flowers, about 100–200 h, as suggested by Sharpe et al. [\[22\]](#page-9-21).

Pollination and fruit set are negatively influenced by high temperatures (>32 \degree C), combined with low relative humidity [\[23\]](#page-9-22). During fruit ripening, a variation of the temperature from day to night of about 10 degrees is important to increase fruit flavor and taste [\[24\]](#page-9-23).

Feijoa has high water requirements, especially during bloom and fruit development. The production of feijoa is favored by precipitation between 700 and 1200 mm/year, but this species tolerates up to about 2000 mm if there is good light and low relative humidity of around 70% [\[23](#page-9-22)[,25\]](#page-10-0). Drought tolerance is different in feijoa genotypes and water-loss of 20–25% in leaves can cause the destruction of photosystem II [\[26\]](#page-10-1).

Stress factors such as drought and high air temperatures during active vegetation in feijoa leaves altered some antioxidant enzyme such as catalase involved in hydrogen peroxide detoxification [\[27\]](#page-10-2).

Feijoa has a low requirement of nitrogen compared to potassium and phosphorus, but an adequate amount of nutrients in every stage of tree growth allows to obtain highyield and high-quality fruit. In young plants (one-year-old), Thorp and Bieleski [\[18\]](#page-9-17) recommended to apply 12–18 kg ha⁻¹ of N, splitting in 3 times, while Morley-Bunker [\[28\]](#page-10-3) suggested to use $25-30$ kg ha⁻¹ of N, probably due to different soil features. Nitrogen fertilization increases gradually until reaching 120 kg ha⁻¹ after ten years of age [\[28\]](#page-10-3). Concerning phosphorus and potassium fertilization, in young plants, the recommended amount is 40 kg ha $^{-1}$ and 20 kg ha $^{-1}$, respectively, that increases gradually up to 80 kg ha $^{-1}$ and 80 kg ha⁻¹ respectively in adult trees [\[28\]](#page-10-3). In a feijoa mature orchard, excessive amounts of nitrogen can cause high fruit development and negatively affects postharvest shelf-life [\[29\]](#page-10-4).

Arioli et al. [\[29\]](#page-10-4), by analyzing fruit composition, determined the amount of nutrients that plants take up for an annual yield of 20 t ha⁻¹, which is: 26 kg (N); 1.5 kg (P); 41 kg (K); 2.4 kg (Ca); 1.9 kg (Mg).

In Italy, feijoa commercial orchards are subject to few pests and diseases compared to Brazil where this species shows a high susceptibility to pathogens [\[30\]](#page-10-5). Anthracnose (*Colletotrichum gloeosporioides*) is the main disease that affects this crop and causes important fruit losses both in the field, while fruit are still on the tree, and during postharvest life [\[19](#page-9-18)[,30\]](#page-10-5). Recently, *C. siamense* has also been identified in feijoa fruit with well-defined dark spots of irregular shape [\[31\]](#page-10-6). *Botrytis cinerea* can cause enormous losses in crops generating rot in unripe fruits during the rain periods [\[32\]](#page-10-7). Other adversities are the basal stem rot and the root rot due to fungi of genus *Phytophthora*, *Pythium,* and *Rosellinia*.

Feijoa is considered a pest-free crop, but occasionally attacks of black scale and fruit flies (*Ceratitis capitata*) have been observed in California [\[33\]](#page-10-8). Presence of leafroller, mealybug, hard wax scale has been reported in New Zealand. These pests do not produce big loss, but can reduce yields by damaging leaves and small shoots [\[34\]](#page-10-9).

3. Genetic Resources and Molecular Tools

Feijoa cultivars widespread in the world are all based on the Uruguayan fruit type with soft juicy flesh and small seeds and classified into three groups based on ripeningtime (early, mid, and late) [\[4\]](#page-9-3). The development of new cultivars with superior fruit quality traits and adapted for new environments is needed to accelerate the commercial exploitation of this species. Genetic variability of this species is maintained in germplasm collections around the world such as the Feijoa Active Germplasm Bank (BAG) in Sao Joaquim-SC (Brazil), the National Feijoa Center (CENAF) in La Vega (Cundinamarca, Colombia), and the Nikitsky Botanical Garden (Crimean Peninsula) that preserve 313, 1500, and 400 accessions, respectively, and develop novel resources for genetics and breeding research [\[11,](#page-9-10)[35\]](#page-10-10). For fruit tree species, such as feijoa, the breeding process is slow and costly because of the long juvenile period, extensive phenotyping cost, and limited field space [\[36\]](#page-10-11). In fact, fruit production starts in year 2–3 after planting, gaining full production in years 5–8 and the length of the juvenile phase seems to be determined by pruning strategies rather than from genotype differences [\[37\]](#page-10-12). For this reason, breeding for new genetic resources is currently based on the use of molecular markers that are also useful for characterization and marketing of existing cultivars. Nonetheless, genomic resources in NCBI for A. sellowiana consist altogether of a total of 197,247 referring to 54 DNA fragments, which also include the complete sequence of the chloroplast (159,370 bp) [\[38\]](#page-10-13). Among these fragments, 24 microsatellites (13,882 bases) are included, predominantly transferred from Eucalyptus to feijoa [\[39](#page-10-14)[,40\]](#page-10-15). The species has been the subject of several sequence studies aimed mainly at establishing taxonomic relationships in the Myrtaceae family [\[41–](#page-10-16)[43\]](#page-10-17). These studies have mainly analyzed the intergenic spaces of the chloroplast, ribosomal genes, maturase K, and NADH dehydrogenase. Sequences obtained in the context of barcoding projects with applications also in the epidemiological field are also worth mentioning [\[44\]](#page-10-18). From a more strictly genetic point of view, it is worth mentioning the in-depth characterization of the leafy gene and its regulation, which is crucial for the induction of flower buds [\[45\]](#page-10-19). Altogether, markers available are limited to few simple sequence repeat (SSR) [\[38](#page-10-13)[,39](#page-10-14)[,46\]](#page-10-20), and to several random amplified polymorphic DNA (RAPD) [\[47\]](#page-10-21) and sequence-related amplified polymorphism (SRAP) [\[6\]](#page-9-5), with a low need for genomic resources. The molecular diversity of the species is documented in few studies in which low-throughput molecular markers are used [\[6](#page-9-5)[,48,](#page-10-22)[49\]](#page-10-23). Amplified fragment length polymorphisms (AFLP), intersimple sequence repeat (ISSR), and a small number of simple sequence repeats (SSR) markers were used in the construction on the first genetic map of the species [\[50\]](#page-10-24). This map represents the first one developed for fruit species within the Myrtaceae family. Recently, a new high-density map has been developed for feijoa based on SNP markers. This was based on new genomic resources that were released in form of SRA databases [\[51\]](#page-11-0). The availability of SRA databases opens the possibility for

the construction of new user-friendly markers based on single nucleotide polymorphism (SNP), useful for breeding and cultivar identification. Markers on SNP based on FRET chemistry, as an example, are emerging as useful and reliable for fruit tree genetic resource management [\[52–](#page-11-1)[54\]](#page-11-2).

4. The Edible Flowers of *A. sellowiana*

The flowers of the feijoa are axillary, erect, actinomorphic, hermaphrodite, showy, very abundant on the plant, and often collected in groups. Four pubescent green glaucous sepals are arranged alternately with respect to the petals form the calyx. The corolla is composed of four to six fleshy petals, pinkish-white inside and white outside, with the edges folded upwards with respect to the major petal axis. The androecium is composed of very showy crimson red stamens with yellow anthers, their number and length varying according to the different varieties. The stigma, also coral-colored, protrudes beyond the stamens and is connected to the inferior ovary with three or four niches. In each niche, there are 36–60 ovules [\[55](#page-11-3)[,56\]](#page-11-4). Feijoa varieties and genotypes differ not only in the shape and size of their fruits, but also in the intensity and hue of the flower color [\[57\]](#page-11-5). Feijoa flowers do not produce nectar and they attract pollinators, mostly birds and insects, with petals rich in sugars [\[58\]](#page-11-6). The sweet taste of the feijoa petals makes them interesting also to enrich the sweet or savory preparations of chefs. The feijoa petals tend to melt on the tongue and become creamy; they are sweet, their taste is reminiscent of papaya and melon with floral notes and a blueberry aftertaste without any hint of acidity. This feature opens a new type of market that requires cultivation measures for the development of a new niche product. Like other ornamental and aromatic plants [\[59](#page-11-7)[–61\]](#page-11-8) whose flowers are edible, the production of flowers must use the organic system without pesticides. In addition to an aesthetic interest, feijoa petals contain secondary compounds of interest to human health. Recent studies [\[62,](#page-11-9)[63\]](#page-11-10) have examined compounds present in feijoa flowers. Among the most present secondary compounds are pedunculagin, gallic acid, cyanidin glucoside (an anthocyanin responsible for the color of the petals), ellagic acid (a phenolic compound), a flavone, and a flavonoid (gossypetin arabinofuranoside). Some of these compounds have pharmacological importance. For example, ellagic acid is known for its anti-inflammatory, antimutagenic, antioxidant, antimicrobial, and anticarcinogenic activity [\[64\]](#page-11-11). The latter activity is also displayed by the flavone, gallic acid, and the pedunculagin. Magri and collaborators [\[7\]](#page-9-6) investigated the bioactive compounds change during flower development and senescence: the petals contain the greatest amount of bioactive compounds when the flowers are not fully open and the filaments and anthers are dark reddish in color (F2 stage). At stage F2, the feijoa petals show peaks in both polyphenols and flavonoids, anthocyanins, and ascorbic acid content, and then at this stage, the antioxidant activity is also higher. Generally, the secondary compounds present in the petals influence their antioxidant activity, and the latter is highly correlated to total polyphenolic compounds in edible flowers [\[60\]](#page-11-12). The decline in bioactive compound content during feijoa flower development is probably due to active degradation or dilution of compounds resulting from the expansion of the petals. The inner color of feijoa petals changes during flower development becoming a good indicator: with senescence, feijoa petals lose their color and the intense pink fades to purple and finally in white. The color loss is accompanied by wilting, making the petals inappropriate for marketing and consumption. Considering the massive flower generation during the bloom, storage methods can be used to avoid the loss of the product part. De Souza et al. [\[65\]](#page-11-13) tested cold postharvest conservation of feijoa flowers, applying different concentration of salicylic acid, ascorbic acid, sucrose, and 1-methylcyclopropene. The authors recorded that the cold storage (10 °C) with 1-methylcyclopropene at 500 μ L L⁻¹ is the best treatment to conserve the petal color, delaying the wilting and the darkening of feijoa petals for at least four days. To date, no trials have been conducted on the drying of the flowers or processed in different preparations (sauces, preserves etc.) In fact, feijoa flowers can be used in formulations for food and pharmaceutical supplements or confectionery industries. Processes and technologies can be applied to improve the feijoa flower preservation but so far, no sufficient trials have been carried out to extend the flower shelf life of feijoa.

5. Fruit: Nutraceutical Compounds and Health Benefits

Feijoa fruits are a source of essential nutrients for human health, such as vitamins, minerals, dietary fibers, and various bioactive compounds, which have been reported to have positive effects on the reduction of incidents of degenerative diseases [\[3,](#page-9-2)[66\]](#page-11-14). Physicochemical characteristics are shown in Table [1.](#page-5-0) Several studies have demonstrated that variations in the fruit weight, size and shape were mostly influenced by the genotype, soil, climatic conditions, and the fruit maturity stage $[6,67–69]$ $[6,67–69]$ $[6,67–69]$. The total acid (TA) content and total soluble solids (TSS) are important factors for fruit quality, while the ratio TSS:TA is usually used for determination of the taste and palatability of the fruit and, therefore, the consumer acceptability [\[69\]](#page-11-16).

Table 1. Physicochemical and nutritional characteristics of fresh feijoa fruit.

The main compounds contributing to soluble solids (TSS) content are fructose, glucose, and sucrose [\[70\]](#page-11-17). At fruit maturity, total sugar content ranged between 10–16 ◦Brix. Most fruits are sweet, have a pleasant texture and flavor, and are eaten as a dessert or snack-food. Titratable acid is mainly dominated by malic and citric acid in a similar concentration [\[70\]](#page-11-17).

Table [1](#page-5-0) shows the nutritional composition of feijoa. Water is the most abundant component and makes up about 85% of the fruit [\[3\]](#page-9-2). The fruit is characterized by a good amount of carbohydrates, low protein content, as well as fat. For these reasons, feijoa fruit is considered a valid diet adjuvant. Feijoa appears to be a good source of dietary fiber (3.8–5.0% of fruit fresh weight) [\[71\]](#page-11-18). It is well documented that an adequate intake of dietary fiber, mainly derived from the peel, is essential for a healthy gut and has also been related to a reduced risk for developing common 'life-style diseases' such as heart disease, certain cancers, and type 2 diabetes [\[69,](#page-11-16)[72\]](#page-11-19).

The fruit is also characterized, like apples, by a very high amount of pectin. It is commonly consumed fresh or as a juice, but it is also used by the food industry to produce some confectionery products, such as sorbet, ice cream, or incorporated into bakery products, sauces, pies, or drinks.

Feijoas are an excellent source of iodine (3 mg/100g FW) [\[73\]](#page-11-20) and, therefore, perform an important preventive action in the treatment of the thyroid gland and in the endocrine system in general [\[71,](#page-11-18)[74\]](#page-11-21). The major mineral is potassium, followed by calcium, magnesium, sodium, iron, and zinc (Table [1\)](#page-5-0). It should be emphasized that the low sodium level, balanced by the high potassium content, allows to maintain the body's salt balance.

These fruits, therefore, could be considered as good sources of vitamin C for the purpose of human nutrition (Table [2\)](#page-6-0) [\[6,](#page-9-5)[69\]](#page-11-16). It is well known that ascorbic acid levels are dependent of a wide variety of environmental factors including light, temperature, salt and drought stress, the presence of atmospheric pollutants, metals, and herbicides, which impact the composition of plant products [\[66\]](#page-11-14).

Table 2. Vitamin C content, total phenolic content, and total antioxidant activity of peel and pulp tissues of the feijoa fruit.

Parameter	Peel	Pulp
Vitamin C (mg ascorbic acid 100 g ⁻¹ FW)	77.64	57.21
Total phenolic content (mg GAE 100 g^{-1} FW)	114.04	85.02
Total antioxidant activity (EC ₅₀ ; mg FW/g ⁻¹ DPPH)	9.04	42.49

In addition to vitamin C, feijoa also contained vitamin E (α -tocopherol). Many studies have shown that a high intake of vitamin E is correlated with a reduced risk of developing non-communicable diseases [\[75\]](#page-11-22). Alpha-tocopherol, the most biologically active form of vitamin E, was found as the main tocopherol constituent in the feijoa fruit samples.

Feijoa contains high amounts of polyphenols (80–120 mg gallic acid/100 g FW) [\[62](#page-11-9)[,76\]](#page-11-23), such as hydroxycinnamic acids (caffeic, chlorogenic, p-coumaric, ferulic and syringic) [\[1\]](#page-9-0), flavonoids (catechin, rutin, quercetin, eriocitrin, eriodictyol, and pyrocatechol) [\[9](#page-9-8)[,62](#page-11-9)[,76\]](#page-11-23) and carotenoids (β-carotene) [\[77](#page-11-24)[,78\]](#page-11-25).

Environmental conditions (temperature, relative humidity, light) and the collection period play a significant role in plant development and influence the biosynthesis of secondary metabolites [\[66\]](#page-11-14).

Numerous studies conducted both in vitro and in vivo have demonstrated the antimicrobial [\[79,](#page-11-26)[80\]](#page-12-0), anticarcinogenic [\[81\]](#page-12-1), anti-ulcer [\[82\]](#page-12-2), antifungal [\[83\]](#page-12-3) and anti-inflammatory [\[84,](#page-12-4)[85\]](#page-12-5) properties of the feijoa plants. The high content of bioactive compounds of these fruits could be a valuable tool to promote their consumption and represent good perspectives for nutraceutical and cosmetic purposes [\[2\]](#page-9-1).

The functional compounds are not only present in the pulp, but also found at a high level in the other tissues such as the leaf and peel. In fact, the edible fruit peel showed higher content of vitamin C and phenolic compounds [\[66\]](#page-11-14) than the pulp (Table [2\)](#page-6-0) with high antioxidant properties [\[1](#page-9-0)[,8\]](#page-9-7). Therefore, there is ample interest in the development technologies to introduce this tissue in nutraceutical products.

Furthermore, feijoa is rich in volatile compounds such as methyl benzoate, ethyl butanoate, and ethyl benzoate [\[3,](#page-9-2)[77\]](#page-11-24), which account for about 90% of the volatile fraction, giving this fruit an 'unique' flavor profile [\[2,](#page-9-1)[86\]](#page-12-6). Limonene (36.2%), β-cariofillene (12%), ledene (9.6%), α-umulene (6.3%), β-elemene (4.9%) and δ-cadinene (4.8%) were the major components in the oil [\[86\]](#page-12-6). Many studies have demonstrated the antitumor and antimicrobial properties of this oil [\[2](#page-9-1)[,69\]](#page-11-16).

The essential oil of *A. sellowiana* may be valuable for the flavoring of foods, such as chewing gums and sweets. Feijoa can be used in other cosmetic products, such as shampoos, soaps, shower gels, as well as in perfumes and deodorants. Furthermore, it could be used in the food preservation due to high antimicrobial compounds [\[80\]](#page-12-0).

6. Postharvest Management

Feijoa is usually consumed as fresh fruit or as juice, but it is also used by the food industry to produce a range of products such as jam, sorbet, ice cream, and fermented beverage [\[78](#page-11-25)[,87\]](#page-12-7). Commercial production of fruits is advanced in New Zealand, Colombia, California, Georgia, and Azerbaijan [\[3,](#page-9-2)[11\]](#page-9-10). In Italy, feijoas are mainly produced for fresh consumption, on a small and local scale, although a slightly increased interest in exporting fruits to reach new market opportunity has been recently observed [\[88\]](#page-12-8). However, the

general low consumer demand seems to be the consequence of a lack of clear information regarding fruits and their nutritional quality [\[89\]](#page-12-9).

Feijoas have a very short harvest season with a limited shelf-life after harvest. Unlike other fruits, feijoas do not change color markedly during ripening, thus it is very difficult to harvest ripe fruit. The technique of 'touch picking', which relies on using low force to detach the fruit from its stalk, is consider a practical and reliable method to harvest ready-to-eat feijoas just prior to the time of fruits' natural drop [\[90\]](#page-12-10). This method is widely accepted as commercial standard in New Zealand, but it is not always suitable for fruit which must withstand long periods in storage, in which early picking is required to reduce chilling injury incidence and retain quality attributes [\[91\]](#page-12-11). The use of near-infrared spectroscopy (NIRS) has high potential as an innovative non-destructive approach to assess the feijoa maturity at harvest [\[92,](#page-12-12)[93\]](#page-12-13). Chlorophyll fluorescence, which indirectly indicates the physiological status of a green tissue, has been studied as a promising nondestructive technique in predicting ripening in the feijoas, but not reliable in detecting chilling injury [\[94\]](#page-12-14).

During the postharvest phase, a variety of changes occur determining the quality of feijoas and leading to fruit senescence [\[95\]](#page-12-15). Although highly variable among cultivars, they generally include weight loss, taste deterioration, a rapid darkening of the flesh, and a higher susceptibility to fungal pathogens [\[96\]](#page-12-16). Anthracnose rot (*Colletotrichum gloeosporioides*) is the main cause of product loss both in the field and after harvest [\[97,](#page-12-17)[98\]](#page-12-18), followed by rots caused by *Botrytis cinerea*, *Penicillium* spp., and *Pestalotiopsis* spp., which particularly occur during the period of storage [\[99](#page-12-19)[,100\]](#page-12-20).

Regarding postharvest handling of fruits, no standard procedure from harvest to final consumption has been documented, and fruit handling specifications are provided differently by markets [\[91\]](#page-12-11). For this reason, in some cases, fruits are brushed to enhance glosses, whereas others are sold with the natural wax, both without washing [\[18\]](#page-9-17).

Various studies have indicated the typical climacteric behavior of fruits with high respiration rates and ethylene production [\[18](#page-9-17)[,101](#page-12-21)[,102\]](#page-12-22); thus, the immediate refrigeration of fruit after harvest is required to reduce respiration rate, transpiration, ethylene activity, and microbial growth [\[96\]](#page-12-16). To eliminate ethylene from storage environments, an adequate ventilation with clean air from outside is necessary. The use of materials able to oxidize or absorb ethylene (zeolite, ozone, potassium permanganate) are recommended when ventilation is not possible, i.e., in controlled atmosphere [\[103\]](#page-12-23). A laser-based photoacoustic detector, with a sensitivity of 0.3 nL L⁻¹, was studied by Gwanpua et al. [\[104\]](#page-12-24) to assess, in real time, fruit physiological status by endogenous ethylene measurement produced.

Feijoa, being a sub-tropical fruit, suffers from chilling injury at the recommended storage temperature of $4 \degree C$. This physiological disorder affects fruits, reducing their marketability. Externally, the peel develops sunken spots that turn from green to brown and finally to black; internally, the vascular elements become brown, and the flesh develops a pink to brown color $[105]$. Often, these symptoms will not be visible until after fruits are transferred to 20 \degree C, making them unsaleable, with the consequence of substantial economic losses [\[18\]](#page-9-17). Although the chilling injury mechanism of development is not still well understood, Sevillano et al. [\[106\]](#page-12-26) noted that ethylene biosynthesis, polyamines (PAs), and cellular abscisic acid (ABA) increased in chilled injury fruit.

Different strategies have been suggested to control chilling injury development in cold stored feijoa. Postharvest application of carnauba-based wax coating resulted in the reduction of weight loss and internal browning [\[107\]](#page-12-27). Treatments with 15% calcium chloride (CaCl₂) solution resulted in rots and weight loss reduction, with beneficial effects on fruits' storage life [\[52\]](#page-11-1). Moreover, the efficacy of intermittent warming was also studies, by alternating shorter warm periods with longer cold period to increase fruit resistance to low temperatures; differently from other fresh products, results showed an accelerated fruit ripening that led to faster deterioration [\[108\]](#page-12-28).

The effect of cold storage on feijoas quality parameters was studied by several re-searchers with the aim to extend fruit shelf-life [\[90,](#page-12-10)[109–](#page-12-29)[111\]](#page-13-0). Application of 5 ± 1 °C and

90–95% relative humidity enabled to achieve four weeks of storage before the organoleptic qualities become altered and the flesh begins to darken. Klein and Thorp [\[100\]](#page-12-20) obtained a varying response about internal browning and fruit quality, on feijoas stored at 4 ◦C for up to four weeks, with a subsequent shelf life of 5–7 days at 20 $^{\circ}$ C, depending on the cultivar, agricultural practices, and maturity. Some cultivars did not show internal browning during the whole storage, whereas others showed browning after only seven days of storage; in most cases, even if the fruit retained an external appearance, the flesh already showed signs of darkening and losses in taste and aroma.

Kwamboka [\[91\]](#page-12-11) found that cold storage by polyliner packaging appeared beneficial to extending feijoa's shelf-life for up to 8 weeks at 1 $°C$, by water loss reduction and firmness and skin color retention. Nonetheless, many fruits developed chilling injury, thereby requiring integrated treatments able to reduce this physiological alteration. Moreover, an increased rots incidence, due to condensation inside packaging, was reported.

In addition to cold storage, controlled atmospheres (CA) and modified atmospheres (MA) have been studied as strategies useful to manipulate the activity of ethylene on fresh produce and extend its storage life. As reported by Zhuang et al. [\[112\]](#page-13-1), inside CA or MAP bags, the high $CO₂$ levels produced inhibited activity of 1-aminocyclopropane-1carboxylate (ACC) synthase, a key enzyme in ethylene biosynthesis. Al-Harthy et al. [\[113\]](#page-13-2) applied CA to refrigerated 'Opal Star' feijoa fruit at $4 °C$, obtaining slightly results in extending shelf-life, by 2.1 oxygen (O_2) and 0% carbon dioxide (CO_2) , while reducing ethylene production, weight loss, and maintaining fruit firmness up to 10 weeks of storage. Despite the good visual fruit appearance, the same authors found that CA affected volatile production by suppression of the main volatile constituent.

Rupavatharam et al. [\[90\]](#page-12-10) noted that the application of the ethylene activity inhibitor 1-methylcyclopropene (1-MCP) to 'Unique' feijoa during storage, resulted in no relevant quality effects on touch picked fruit. On the contrary, fruit harvested 2 weeks prior to touch picked maturity appeared visually acceptable after 6 weeks of storage; however, low soluble solid content (SSC) and high titratable acidity (TA) were reported. Amarante et al. [\[114\]](#page-13-3) reported that the application of 1-MCP at 1500 nL L⁻¹ for 8 h retain firmness and skin color during storage of the cultivar 'Brazilian' for 30 days.

Recent research interest has been focused on different use of feijoas by the development of innovative nanocomposite films for food packaging, made by biodegradable polymeric matrix and natural antioxidant. Through the studies carried out in the last few years, polyethylene oxide (PEO) nanocomposite films were produced, integrating feijoas aqueous extract from waste by-product, bringing an additional use of this multipurpose product as an alternative material to produce active food packaging [\[115\]](#page-13-4). Nanocapsules produced with feijoa and fortified with vitamin D3 were instead studied for the controlled release of active compounds (food fortification), to improve their bioavailability during food processing and storage, thus reducing nutritional deficiencies [\[116\]](#page-13-5).

7. Conclusions and Future Perspectives

Feijoa trees are currently extensively cultivated in California and mainly in New Zealand, but it has a good adaptability to different environments. Climate changes are shifting the established pattern of fruit crops in Mediterranean area, favoring in some areas the cultivations of new tropical fruits. This species, firstly introduced as a home-garden plant, nowadays has begun to be cultivated in commercial orchards for its aromatic and nutraceutical fruits. Furthermore, feijoa edible flowers could be a profitable enterprise niche, especially for farms marketing to consumers and chefs interested in a novel use of edible flowers with high health and nutritional values. Optimization of appropriate strategies and agronomic management are needed to guarantee the sustainability of this fruit crop from farm to table. Furthermore, other studies are needed to advance the development and application of postharvest technologies that facilitate appropriate feijoa management, storage, and a possible re-use of processing waste, in view of a successful transition to a circular economy.

Author Contributions: Writing—original draft preparation, T.V., M.A., A.C., M.G., A.N., M.C.S., and M.P. Writing—review and editing, T.V., M.A., A.C., M.G., A.N., M.C.S. and M.P.; supervision, M.P. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Tuncel, N.B.; Yılmaz, N. Optimizing the extraction of phenolics and antioxidants from feijoa (*Feijoa sellowiana*, Myrtaceae). *J. Food Sci. Technol.* **2015**, *52*, 141–150. [\[CrossRef\]](http://doi.org/10.1007/s13197-013-0968-0)
- 2. Weston, R.J. Bioactive products from fruit of the feijoa (*Feijoa sellowiana*, Myrtaceae): A review. *Food Chem.* **2010**, *121*, 923–926. [\[CrossRef\]](http://doi.org/10.1016/j.foodchem.2010.01.047)
- 3. Zhu, F. Chemical and biological properties of feijoa (*Acca sellowiana*). *Trends Food Sci. Technol.* **2018**, *81*, 121–131. [\[CrossRef\]](http://doi.org/10.1016/j.tifs.2018.09.008)
- 4. Schotsmans, W.C.; Pain, A.S.; Thorp, G. *Feijoa (Acca sellowiana [Berg] Burret)*; Woodhead Publishing Limited: Sawston, UK, 2011; Chapter 6; pp. 115–133.
- 5. de Oliveira Schmidt, H.; Rockett, F.C.; Ebert, G.; Sartori, G.V.; Rezzadori, K.; Rodrigues, R.C.; de Oliveira Rios, A.; Manfroi, V. Effect of enzymatic treatments and microfiltration on the physicochemical quality parameters of feijoa (*Acca sellowiana*) juice. *Int. J. Food Sci. Technol.* **2021**, *56*, 4983–4994. [\[CrossRef\]](http://doi.org/10.1111/ijfs.15223)
- 6. Pasquariello, M.S.; Mastrobuoni, F.; Di Patre, D.; Zampella, L.; Capuano, L.R.; Scortichini, M.; Petriccione, M. Agronomic, nutraceutical and molecular variability of feijoa (*Acca sellowiana* (O. Berg) Burret) germplasm. *Sci. Hortic.* **2015**, *191*, 1–9. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2015.04.036)
- 7. Magri, A.; Adiletta, G.; Petriccione, M. Evaluation of antioxidant systems and ascorbate-glutathione cycle in feijoa edible flowers at different flowering stages. *Foods* **2020**, *9*, 95. [\[CrossRef\]](http://doi.org/10.3390/foods9010095)
- 8. Amarante, C.V.T.; Souza, A.G.; Beninca, T.D.T.; Steffens, C.A. Phenolic content and antioxidant activity of fruit of Brazilian genotypes of feijoa. *Pesqui. Agropecu. Bras.* **2017**, *52*, 1223–1230. [\[CrossRef\]](http://doi.org/10.1590/s0100-204x2017001200011)
- 9. Sun-Waterhouse, D.; Wang, W.; Waterhouse, G.I.N.; Wadhwa, S.S. Utilisation potential of feijoa fruit wastes as ingredients for functional foods. *Food Bioprocess Technol.* **2013**, *6*, 3441–3455. [\[CrossRef\]](http://doi.org/10.1007/s11947-012-0978-3)
- 10. Campos, D.A.; Gómez-García, R.; Vilas-Boas, A.A.; Madureira, A.R.; Pintado, M.M. Management of fruit industrial by-products-A case study on circular economy approach. *Molecules* **2020**, *25*, 320. [\[CrossRef\]](http://doi.org/10.3390/molecules25020320)
- 11. Fischer, G.; Parra-Coronado, A. Influence of some environmental factors on the feijoa (*Acca sellowiana* [Berg] Burret): A review. *Agron. Colomb.* **2020**, *38*, 388–397. [\[CrossRef\]](http://doi.org/10.15446/agron.colomb.v38n3.88982)
- 12. Pugliano, G. La feijoa sellowiana. Principali caratteristiche colturali e prospettive per l'immediato futuro. *Spec. Fruttic. Esotica* **1987**, *69–70*, 13–20.
- 13. Ramirez, F.; Kallarackal, J. Feijoa [*Acca sellowiana* (O. Berg) Burret] pollination: A review. *Sci. Hortic.* **2017**, *226*, 333–341. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2017.08.054)
- 14. Zou, F.; Chen, S.L.; Yuan, D.Y.; Zhang, R.Q.; Zhang, L.; Xiong, H. Microsporogenesis, megasporogensis and male and female gametophyte development in *Feijoa sellowiana* (Myrtaceae). *Int. J. Agric. Biol.* **2016**, *18*, 637–642. [\[CrossRef\]](http://doi.org/10.17957/IJAB/15.0140)
- 15. Finatto, T.; Dos Santos, K.L.; Steiner, N.; Bizzocchi, L.; Holderbaum, D.F.; Ducroquet, J.P.H.J.; Guerra, M.P.; Nodari, R.O. Late-acting self-incompatibility in *Acca sellowiana* (Myrtaceae). *Aust. J. Bot.* **2011**, *59*, 53–60. [\[CrossRef\]](http://doi.org/10.1071/BT10152)
- 16. Esemann-Quadros, K.; Mota, A.P.; Kerbauy, G.B.; Guerra, M.P.; Ducroquet, J.P.H.J.; Pescador, R. Estudio anatômico do crescimento do fruto em *Acca sellowiana* Berg. *Rev. Bras. Frutic.* **2008**, *30*, 296–302. [\[CrossRef\]](http://doi.org/10.1590/S0100-29452008000200005)
- 17. Quintero, O.C. Feijoa (*Acca sellowiana* Berg). In *Manual Para el Cultivo de Frutales en el Trópico*; Fischer, G., Ed.; Produmedios: Bogotá, Colombia, 2012; pp. 443–473.
- 18. Thorp, T.G.; Bieleski, R.L. *Feijoas: Origins, Cultivation and Uses*; David Bateman Publishing: Auckland, New Zealand, 2002; p. 87.
- 19. Fischer, G.; Miranda, D.; Cayón, G.; Mazorra, M. *Cultivo, Poscosecha y Exportación de la Feijoa (Acca sellowiana Berg)*; Produmedios: Bogotá, Colombia, 2003; p. 152.
- 20. Quintero-Castillo, O. Selección de cultivares, manejo del cultivo y regulación de cosechas de feijoa. In *Cultivo, Poscosecha y Exportación de la Feijoa (Acca sellowiana Berg)*; Fischer, G., Miranda, D., Cayón, G., Mazorra, M., Eds.; Produmedios: Bogotá, Colombia, 2003; pp. 49–71.
- 21. Silva, L.R.; Moura, A.P.C.; Gil, B.; Rohr, A.; Almeida, S.M.Z.; Donazzolo, J.; Perboni, A.T.; Oliveira, F.L.R.; Sant'Anna-Sontos, B.F.; GAlon, L.; et al. Morphophysiological changes of *Acca sellowiana* (Myrtaceae: Myrtoideae) saplings under shade gradient. *Braz. J. Biol.* **2021**, *84*, e252364. [\[CrossRef\]](http://doi.org/10.1590/1519-6984.252364) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/35019092)
- 22. Sharpe, R.H.; Sherman, W.B.; Miller, E.P. Feijoa, history and improvement. *Sel. Proc. Fla. State Hortic. Soc.* **1993**, *106*, 134–139.
- 23. Duarte, O.; Paull, R. *Exotic Fruits and Nuts of the New World*; CABI: Wallingford, UK, 2015.
- 24. Taiz, L.; Zeiger, E. *Fisiologia Vegetal*, 5th ed.; Artemed: Porto Alegre, Brazil, 2013.
- 25. Pachón, G.; Quintero, O. La feijoa (*Feijoa sellowiana* Berg.) fruta promisoria para Colombia. *Acta Hortic.* **1992**, *310*, 239–248. [\[CrossRef\]](http://doi.org/10.17660/ActaHortic.1992.310.29)
- 26. Shishkina, E.; Gubanova, T.; Titov, V. Some special features of the water regime and the photosynthetic apparatus activity in *Feijoa sellowiana* (o. Berg) O. Berg plants under the water stress. *BIO Web Conf.* **2021**, *38*, 00117. [\[CrossRef\]](http://doi.org/10.1051/bioconf/20213800117)
- 27. Omarova, Z.; Platonova, N.; Belous, O.; Omarov, M. Evaluation of the physiological state of feijoa (*Feijoa sellowiana* Berg) in subtropical Russia. *Slovak J. Food Sci.* **2020**, *14*, 286–291. [\[CrossRef\]](http://doi.org/10.5219/1290)
- 28. Morley-Bunker, M. Feijoas. In *Temperate and Subtropical Fruit Production*, 2nd ed.; Jackson, D.I., Looney, N.E., Eds.; CABI Publishing: Cambridge, MA, USA, 1999; pp. 267–269.
- 29. Arioli, C.J.; Moretti Ferreira Pinto, F.A.; Araùjo, L.; Ciotta, M.N.; da Silveira Pasa, M. *A Cultura da Goiabeira-Serrana*; Epagri: Florianópolis, Brazil, 2018.
- 30. Araújo, L.; Pinto, F.A.M.F. Principais doenças e seu controle. In *A Cultura da Goiabeira-Serrana*; Ciotta, M.N., Arioli, C.J., Pinto, F.A.M.F., Santos, K., Araújo, L., Pasa, M.S., Eds.; Epagri: Florianópolis, Brazil, 2018; pp. 147–166.
- 31. Fantinel, V.S.; Muniz, M.F.B.; Blume, E.; Araújo, M.M.; Poletto, T.; Dutra, A.; Tonetto, T.; Harakava, R.; Maciel, C.G. First Report of *Colletotrichum siamense* causing anthracnose on *Acca sellowiana* fruits in Brazil. *Plant Dis.* **2017**, *101*, 1035. [\[CrossRef\]](http://doi.org/10.1094/PDIS-01-17-0096-PDN)
- 32. Perea Dallos, M.; Fischer, G.; Miranda, D. *Feijoa Acca sellowiana [Berg] Burret*; Chapter-January 2010; Research Gate: Berlin, Germany, 2010.
- 33. Segura, D.F.; Vera, M.T.; Cagnotti, C.L.; Vaccaro, N.; Decoll, O.R.; Ovruski, S.M.; Cladera, J.L. Relative abundance of *Ceratitis capitata* and *Anastrepha fraterculus* (Diptera: Tephritidae) in diverse host species and localities of Argentina. *Ann. Entomol. Soc. America* **2006**, *99*, 70–83. [\[CrossRef\]](http://doi.org/10.1603/0013-8746(2006)099[0070:RAOCCA]2.0.CO;2)
- 34. Janick, J.; Paull, R.E. *The Encyclopedia of Fruit and Nuts*, 1st ed.; CABI: Wallingford, UK, 2007; pp. 526–532.
- 35. Sánchez-Mora, F.D.; Saifert, L.; Ciotta, M.N.; Ribeiro, H.; Petry, V.S.; Rojas-Molina, A.; Lopes, M.E.; Lombardi, G.; Santos, K.; Ducroquet, J.; et al. Characterization of phenotypic diversity of feijoa fruits of germplasm accessions in Brazil. *Agrosyst. Geosci. Environ.* **2019**, *2*, 190005. [\[CrossRef\]](http://doi.org/10.2134/age2019.01.0005)
- 36. Byrne, D.H. Trends in fruit breeding. In *Fruit Breeding*; Badenes, M.L., Byrne, D.H., Eds.; Springer: New York, NY, USA, 2012; pp. 3–36.
- 37. Li, J.K.; Yang, J.H.; Liu, Y.J.; Huang, J.X.; Wu, C.X.; Liu, T.; Gong, W.Q. Growth and Development of Feijoa with Different Plant Shape and in Greenhouses Environment. In *Advanced Materials Research*; Trans Tech Publications, Ltd.: Freienbach, Switzerland, 2014; Volume 886, pp. 294–298. [\[CrossRef\]](http://doi.org/10.4028/www.scientific.net/amr.886.294)
- 38. de Oliveira Machado, L.; do Nascimento Vieira, L.; Stefenon, V.M.; de Oliveira Pedrosa, F.; de Souza, E.M.; Guerra, M.P.; Nodari, R.O. Phylogenomic relationship of feijoa (*Acca sellowiana* (O.Berg) Burret) with other Myrtaceae based on complete chloroplast genome sequences. *Genetica* **2017**, *145*, 163–174. [\[CrossRef\]](http://doi.org/10.1007/s10709-017-9954-1) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/28185042)
- 39. dos Santos, K.L.; Welter, L.J.; Dantas, A.C.M.; Guerra, M.P.; Ducroquet, J.P.H.J.; Nodari, R.O. Transference of microsatellite markers from *Eucalyptus* spp. to *Acca sellowiana* and the successful use of this technique in genetic characterization. *Genet. Mol. Biol.* **2007**, *30*, 73–79. [\[CrossRef\]](http://doi.org/10.1590/S1415-47572007000100014)
- 40. dos Santos, K.L.; Santos, M.O.; Laborda, P.R.; Souza, A.P.; Peroni, N.; Nodari, R.O. Isolation and characterization of microsatellite markers in *Acca sellowiana* (Berg) Burret. *Mol. Ecol. Resour.* **2008**, *8*, 998–1000. [\[CrossRef\]](http://doi.org/10.1111/j.1755-0998.2008.02134.x) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/21585953)
- 41. Mazine, F.; Quintino Faria, J.E.; Giaretta, A.; Vasconcelos, T.; Forest, F.; Lucas, E. Phylogeny and biogeography of the hyper– diverse genus *Eugenia* (Myrtaceae: Myrteae), with emphasis on E. sect. Umbellatae, the most unmanageable clade. *Taxon* **2018**, *67*, 752–769. [\[CrossRef\]](http://doi.org/10.12705/674.5)
- 42. Vasconcelos, T.N.C.; Proença, C.E.B.; Ahmad, B.; Aguilar, D.S.; Aguilar, R.; Amorim, B.S.; Campbell, K.; Costa, I.R.; De-Carvalho, P.S.; Faria, J.E.Q.; et al. Myrteae phylogeny, calibration, biogeography and diversification patterns: Increased understanding in the most species rich tribe of Myrtaceae. *Mol. Phylogenet. Evol.* **2017**, *109*, 113–137. [\[CrossRef\]](http://doi.org/10.1016/j.ympev.2017.01.002)
- 43. Lucas, E.J.; Harris, S.A.; Mazine, F.F.; Belsham, S.R.; Lughadha, E.M.N.; Telford, A.; Gasson, P.E.; Chase, M.W. Suprageneric phylogenetics of Myrteae, the generically richest tribe in Myrtaceae (Myrtales). *Taxon* **2007**, *56*, 1105–1128. [\[CrossRef\]](http://doi.org/10.2307/25065906)
- 44. Buys, M.H.; Flint, H.J.; Miller, E.M.; Yao, H.; Caird, A.R.; Ganley, R.J. Preparing for the invasion: Efficacy of DNA barcoding to discern the host range of myrtle rust (*Puccinia psidii*) among species of Myrtaceae. *Forestry* **2016**, *89*, 263–270. [\[CrossRef\]](http://doi.org/10.1093/forestry/cpw017)
- 45. Feng, Y.Z.; Zhang, L.; Yuan, D.Y.; Long, H.X.; Liu, M.; Zhang, R.Q.; Chen, S.L. Expression pattern of FsLFY gene and its promoter cloning in *Feijoa sellowiana*. *J. Plant Genet. Resour.* **2014**, *15*, 831–837.
- 46. Klabunde, G.H.F.; Olkoski, D.; Vilperte, V.; Zucchi, M.I.; Nodari, R.O. Characterization of 10 new nuclear microsatellite markers in *Acca sellowiana* (Myrtaceae). *Appl. Plant Sci.* **2014**, *2*, 1400020. [\[CrossRef\]](http://doi.org/10.3732/apps.1400020)
- 47. Dettori, M.T.; Palombi, M.A. Identification of *Feijoa sellowiana* Berg accessions by RAPD markers. *Sci. Hortic.* **2000**, *86*, 279–290. [\[CrossRef\]](http://doi.org/10.1016/S0304-4238(00)00157-6)
- 48. Donazzolo, J.; Stefenon, V.M.; Guerra, M.P.; Nodari, R.O. On farm management of *Acca sellowiana* (Myrtaceae) as a strategy for conservation of species genetic diversity. *Sci. Hortic.* **2020**, *259*, 108826. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2019.108826)
- 49. Saifert, L.; Sánchez-Mora, F.D.; Borsuk, L.J.; Donazzolo, J.; da Costa, N.C.F.; Ribeiro, H.N.; Nodari, R.O. Evaluation of the genetic diversity in the feijoa accessions maintained at Santa Catarina, Brazil. *Crop Sci.* **2020**, *60*, 345–356. [\[CrossRef\]](http://doi.org/10.1002/csc2.20088)
- 50. Quezada, M.; Pastina, M.M.; Ravest, G.; Silva, P.; Vignale, B.; Cabrera, D.; Hinrichsen, P.; Garcia, A.A.F.; Pritsch, C. A first genetic map of Acca sellowiana based on ISSR, AFLP and SSR markers. *Sci. Hortic.* **2014**, *169*, 138–146. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2014.02.009)
- 51. Quezada, M.; Amadeu, R.R.; Vignale, B.; Cabrera, D.; Pritsch, C.; Garcia, A. Construction of a high-density genetic map of *Acca sellowiana* (Berg.) burret, an outcrossing species, based on two connected mapping populations. *Front. Plant Sci.* **2021**, *12*, 626811. [\[CrossRef\]](http://doi.org/10.3389/fpls.2021.626811)
- 52. Nunziata, A.; Ruggieri, V.; Petriccione, M.; De Masi, L. Single nucleotide polymorphisms as practical molecular tools to support European chestnut agrobiodiversity management. *Int. J. Mol. Sci.* **2020**, *21*, 4805. [\[CrossRef\]](http://doi.org/10.3390/ijms21134805)
- 53. Bernard, A.; Marrano, A.; Donkpegan, A.; Brown, P.J.; Leslie, C.A.; Neale, D.B.; Dirlewanger, E. Association and linkage mapping to unravel genetic architecture of phenological traits and lateral bearing in Persian walnut *(Juglans regia* L.). *BMC Genom.* **2020**, *21*, 203. [\[CrossRef\]](http://doi.org/10.1186/s12864-020-6616-y)
- 54. Zurn, J.D.; Driskill, M.; Jung, S.; Main, D.; Yin, M.H.; Clark, M.C.; Cheng, L.; Ashrafi, H.; Aryal, R.; Clark, J.R.; et al. A Rosaceae family-level approach to identify loci influencing soluble solids content in blackberry for DNA-informed breeding. *G3 Genes Genomes Genet.* **2020**, *10*, 3729–3740. [\[CrossRef\]](http://doi.org/10.1534/g3.120.401449)
- 55. Landrum, L.R. *Camjonesia, Pimenta, Blepharocalix, Legrandia, Acca, Myrrhynium, and Luma (Myrtaceae)*; Fl. Neotrop Monogr 45; New York Botanical Garden: New York, NY, USA, 1986.
- 56. Ramírez, J.M.; Galvis, J.A.; Fischer, G. Postharvest ripening of feijoa (*Acca sellowiana* Berg) treated with CaCl2 at three storage temperatures. *Agron. Colomb.* **2005**, *23*, 117–127.
- 57. Amarante, C.V.T.; do Souza, A.G.; de Benincá, T.D.T.; Steffens, C.A.; Ciotta, M.N. Physicochemical attributes and functional properties of flowers of Brazilian feijoa genotypes. *Pesqui. Agropecu. Bras.* **2019**, *54*, e00445. [\[CrossRef\]](http://doi.org/10.1590/s1678-3921.pab2019.v54.00445)
- 58. Lim, T.K. Acca sellowiana. In *Edible Medicinal and Non Medicinal Plants*; Springer: Dordrecht, The Netherlands, 2012; Volume 3, pp. 601–608.
- 59. Najar, B.; Marchioni, I.; Ruffoni, B.; Copetta, A.; Pistelli, L.; Pistelli, L. Volatilomic analysis of four edible flowers from agastache genus. *Molecules* **2019**, *24*, 4480. [\[CrossRef\]](http://doi.org/10.3390/molecules24244480)
- 60. Marchioni, I.; Pistelli, L.; Ferri, B.; Copetta, A.; Ruffoni, B.; Pistelli, L.; Najar, B. Phytonutritional content and aroma profile changes during postharvest storage of edible flowers. *Front. Plant Sci.* **2020**, *11*, 590968. [\[CrossRef\]](http://doi.org/10.3389/fpls.2020.590968)
- 61. Marchioni, I.; Najar, B.; Ruffoni, B.; Copetta, A.; Pistelli, L.; Pistelli, L. Bioactive compounds and aroma profile of some *Lamiaceae* edible flowers. *Plants* **2020**, *9*, 691. [\[CrossRef\]](http://doi.org/10.3390/plants9060691)
- 62. Aoyama, H.; Sakagami, H.; Hatano, T. Three new flavonoids, proanthocyanidin, and accompanying phenolic constituents from *Feijoa sellowiana*. *Biosci. Biotechnol. Biochem.* **2018**, *82*, 31–41. [\[CrossRef\]](http://doi.org/10.1080/09168451.2017.1412246)
- 63. Montoro, P.; Serreli, G.; Gil, K.A.; D'Urso, G.; Kowalczyk, A.; Tuberoso, C.I.G. Evaluation of bioactive compounds and antioxidant capacity of edible feijoa (*Acca sellowiana* (O. Berg) Burret) flower extracts. *J. Food Sci. Technol.* **2020**, *57*, 2051–2060. [\[CrossRef\]](http://doi.org/10.1007/s13197-020-04239-2) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/32431331)
- 64. Vattem, D.A.; Shetty, K. Biological functionally of ellagic acid: A review. *J. Food Biochem.* **2005**, *29*, 234–266. [\[CrossRef\]](http://doi.org/10.1111/j.1745-4514.2005.00031.x)
- 65. de Souza, A.G.D.; Amarante, C.V.T.; Steffens, C.A.; Beninca, T.D.T.; Padilha, M. Postharvest quality of feijoa flowers treated with different preservative solutions and 1-methylcyclopropene. *Rev. Bras. Frutic.* **2016**, *38*, e-759. [\[CrossRef\]](http://doi.org/10.1590/0100-29452016759)
- 66. Monforte, M.T.; Lanuzza, F.; Mondello, F.; Naccari, C.; Pergolizzi, S.; Galati, E.M. Phytochemical composition and gastroprotective effect of *Feijoa sellowiana* Berg fruits from Sicily. *J. Coast Life Med.* **2014**, *2*, 14–21.
- 67. Sestras, R.; Tamas, E.; Sestras, A. Morphological and genetic peculiarities of fruits in several winter apple varieties with confer resistance to damage. *Agron. Res.* **2006**, *4*, 55–62.
- 68. Usenik, V.; Fajt, N.; Mikulic-Petkovsek, M.; Slatner, A.; Stampar, F.; Veberic, R. Sweet cherry pomological and biochemical characteristics influenced by rootstock. *J. Agric. Food Chem.* **2010**, *58*, 4928–4933. [\[CrossRef\]](http://doi.org/10.1021/jf903755b) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/20337477)
- 69. Phan, A.D.T.; Chaliha, M.; Sultanbawa, Y.; Netzel, M.E. Nutritional characteristics and antimicrobial activity of Australian grown feijoa (*Acca sellowiana*). *Foods* **2019**, *8*, 376. [\[CrossRef\]](http://doi.org/10.3390/foods8090376) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31480592)
- 70. Harman, J.E. Feiioa fruit: Growth and chemical composition during development. *N. Z. J. Agric. Res.* **1987**, *15*, 209–215. [\[CrossRef\]](http://doi.org/10.1080/03015521.1987.10425561)
- 71. Romero-Rodriguez, M.; Vazquez-Oderiz, M.; Lopez-Hernandez, J.; Simal-Lozano, J. Composition of babaco, feijoa, passionfruit and tamarillo produced in Galicia (North-west Spain). *Food Chem.* **1994**, *94*, 23–27. [\[CrossRef\]](http://doi.org/10.1016/0308-8146(94)90227-5)
- 72. Sun-Waterhouse, D.; Teoh, A.; Massarotto, C.; Wibisono, R.; Wadhwa, S. Comparative analysis of fruit-based functional snack bars. *Food Chem.* **2010**, *119*, 1369–1379. [\[CrossRef\]](http://doi.org/10.1016/j.foodchem.2009.09.016)
- 73. Ferrara, L.; Montesano, D. Nutritional characteristics of *Feijoa sellowiana* fruit: The iodine content. *Riv. Sci. Dell'alimentazione* **2001**, *30*, 353–356.
- 74. Migliuolo, G.; Ruggeri, P. Quantitative determination of organic iodine content in *Feijoa sellowiana*. *Riv. Merceol.* **1994**, *33*, 29–36.
- 75. Raiola, A.; Tenore, G.C.; Barone, A.; Frusciante, L.; Rigano, M.M. Vitamin E content and composition in tomato fruits: Beneficial roles and bio-fortification. *Int. J. Mol. Sci.* **2015**, *16*, 29250–29264. [\[CrossRef\]](http://doi.org/10.3390/ijms161226163)
- 76. Peng, Y.; Bishop, K.S.; Quek, S.Y. Extraction optimization, antioxidant capacity and phenolic profiling of extracts from flesh, peel and whole fruit of New Zealand grown feijoa cultivars. *Antioxydant* **2019**, *8*, 141. [\[CrossRef\]](http://doi.org/10.3390/antiox8050141) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31117250)
- 77. Ruberto, G.; Tringali, C. Secondary metabolites from the leaves of *Feijoa sellowiana* Berg. *Phytochemistry* **2004**, *65*, 2947–2951. [\[CrossRef\]](http://doi.org/10.1016/j.phytochem.2004.06.038)
- 78. Sartori, G.V.; Montibeller, M.J.; Furini, G.; Veeck, A.P.D.L.; Sganzerla, W.G.; Beling, P.C.; Rios, A.D.O.; Manfroi, V. Characterization of feijoa fermented beverage. *Int. J. Agric. Nat. Resour.* **2020**, *7*, 35–45. [\[CrossRef\]](http://doi.org/10.7764/ijanr.v47i1.1939)
- 79. Motohashi, N.; Kawase, M.; Shirataki, Y.; Tani, S.; Saito, S.; Sakagami, H. Biological activity of *Feijoa* peel extracts. *Anticancer. Res.* **2009**, *20*, 4323–4329.
- 80. Elfarnini, M.; Abdel-hamid, A.A.; Achir, M.; Jamaleddine, J.; Blaghen, M. Volatile compounds in the skin essential oil of Moroccan *Feijoa sellowiana*. *Eur. J. Med. Plants* **2018**, *23*, 1–7.
- 81. Bontempo, P.; Mita, L.; Miceli, M.; Doto, A.; Nebbioso, A.; De Bellis, F. *Feijoa sellowiana* derived natural flavone exerts anti-cancer action displaying HDAC inhibitory activities. *Int. J. Biochem. Cell Biol.* **2007**, *39*, 1902–1914. [\[CrossRef\]](http://doi.org/10.1016/j.biocel.2007.05.010)
- 82. Leuzzi, A.; Galati, E.M.; Mondello, M.R.; Monforte, M.T. Antiulcer activity of *Feijoa sellowiana* (Myrtaceae): Morphological study. *Planta Med.* **2009**, *75*, 1055.
- 83. Vuotto, M.L.; Basile, A.; Moscatiello, V.; De Sole, P.; Castaldo-Cobianchi, R.; Laghi, E.; Ielpo, M.T.L. Antimicrobial and antioxidant activities of *Feijoa sellowiana* fruit. *Int. J. Antim. Ag.* **2000**, *13*, 197–201.
- 84. Isobe, Y.; Kase, Y.; Narita, M.; Komiya, T. Antioxidative activity of a polyphenol extract from *Feijoa sellowiana* Berg. and its application to cookies. *Nippon Kasei Gakkaishi* **2004**, *55*, 799–804.
- 85. Rossi, A.; Rigano, D.; Pergola, C.; Formisano, C.; Basile, A.; Bramanti, P. Inhibition of inducible nitric oxide synthase expression by an acetonic extract from *Feijoa sellowiana* Berg. fruits. *J. Agric. Food Chem.* **2007**, *55*, 5053–5061.
- 86. Fernandez, X.; Loiseau, A.M.; Poulain, S.; Lizzani-Cuvelier, L.; Monnier, Y. Chemical composition of the essential oil from feijoa (*Feijoa sellowiana* Berg). *peel. J. Essent. Oil Res.* **2004**, *16*, 274–275. [\[CrossRef\]](http://doi.org/10.1080/10412905.2004.9698719)
- 87. Smeriglio, A.; Denaro, M.; De Francesco, C.; Cornara, L.; Barreca, D.; Bellocco, E.; Ginestra, G.; Mandalari, G.; Trombetta, D. Feijoa fruit peel: Micro-morphological features, evaluation of phytochemical profile, and biological properties of its essential oil. *Antioxidants* **2019**, *8*, 320. [\[CrossRef\]](http://doi.org/10.3390/antiox8080320) [\[PubMed\]](http://www.ncbi.nlm.nih.gov/pubmed/31430937)
- 88. Vatrano, T.; Amenta, M.; Copetta, A.; Guardo, M.; Nunziata, A.; Strano, M.C.; Petriccione, M. Feijoa, cresce l'interesse per un frutto esotico dalle molte proprietà. *Riv. Fruttic. Ortofloric.* **2020**, *8*, 48–52.
- 89. Kabiri, S.; Gheibi, F.; Joker, M.; Basiri, S. Antioxidant activity and physicochemical properties of fresh, dried and infused herbal extract of Feijoa Fruit. *Nat. Sci.* **2016**, *14*, 64–70. [\[CrossRef\]](http://doi.org/10.7537/marsnsj141216.12)
- 90. Rupavatharam, S.; East, A.R.; Heyes, J.A. Re-evaluation of harvest timing in 'Unique'feijoa using 1-MCP and exogenous ethylene treatments. *Postharvest Biol. Technol.* **2015**, *99*, 152–159. [\[CrossRef\]](http://doi.org/10.1016/j.postharvbio.2014.08.011)
- 91. Kwamboka, O.J. An Examination of Postharvest Techniques to Enable Sea Freight Export of Feijoa (*Acca sellowiana* [O.Berg.] Burret). Ph.D. Thesis, Massey University, Palmerston North, New Zealand, 2020.
- 92. Olarewajua, O.O.; Bertling, I.; Magwaza, L.S. Non-destructive evaluation of avocado fruit maturity using near infrared spectroscopy and PLS regression models. *Sci. Hortic.* **2016**, *199*, 229–236. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2015.12.047)
- 93. Li, B.; Lecourt, J.; Bishop, G. Advances in non-destructive early assessment of fruit ripeness towards defining optimal time of harvest and yield prediction—A review. *Plants* **2018**, *7*, 3. [\[CrossRef\]](http://doi.org/10.3390/plants7010003)
- 94. Oseko, J.; East, A.; Heyes, J. Can changes in chlorophyll fluorescence be used to determine chilling injury of cold stored feijoa? *Acta Hortic.* **2020**, *1275*, 125–132. [\[CrossRef\]](http://doi.org/10.17660/ActaHortic.2020.1275.18)
- 95. Oseko, J.; East, A.; Heyes, J. Recent advances in the postharvest technology of feijoa. *Sci. Hortic.* **2022**, *297*, 110969. [\[CrossRef\]](http://doi.org/10.1016/j.scienta.2022.110969)
- 96. Amarante, C.V.T.D.; de Souza, A.G.; Benincá, T.D.T.; Steffens, C.A. Fruit quality of Brazilian genotypes of feijoa at harvest and after storage. *Pesqui. Agropecu. Bras.* **2017**, *52*, 734–742. [\[CrossRef\]](http://doi.org/10.1590/s0100-204x2017000900005)
- 97. Itako, A.T.; Tolentino, J.J.B.; Pinto Raduan, J.L.F.; do Prado Mattos, A.; dos Santos, K.L.; Ciotta, M.R. Effect of essential oils on the development of *Colletotrichum* sp. fungus in fragments of *Feijoa sellowiana* fruits. *Acta Sci. Biol. Sci.* **2021**, *43*, e53055. [\[CrossRef\]](http://doi.org/10.4025/actascibiolsci.v43i1.53055)
- 98. Fantinel, V.S.; Muniz, M.F.B.; Poletto, T.; Harakava, R.; Ciotta, M.N.; Savian, L.G.; Favaretto, R.F.; Krahn, J.R.T. Pathogenicity and susceptibility/resistance reaction of feijoa (*Feijoa sellowiana*) cultivars to anthracnose. *Rev. Bras. Ciên. Agrárias* **2020**, *15*, e8395. [\[CrossRef\]](http://doi.org/10.5039/agraria.v15i3a8395)
- 99. Klein, J.D.; Thorp, T.G. Feijoas: Post-harvest handling and storage of fruit. *N.Z. J. Exp. Agric.* **1987**, *15*, 217–221. [\[CrossRef\]](http://doi.org/10.1080/03015521.1987.10425562)
- 100. Minischetti, P.; Porta-Puglia, A.; Cacioppo, O. Fruit rot of *Feijoa sellowiana* caused by *Pestalotiopsis psidii*. *Inf. Fitopatol.* **1991**, *41*, 61–63.
- 101. Parra, A.C.; Fischer, G. Ripening and postharvest behavior in the pineapple guava (*Acca sellowiana* (O. Berg) Burret). A review. *Rev. Colomb. Cienc. Hortic.* **2013**, *7*, 98–110.
- 102. Wills, R.; Golding, J. *Postharvest: An Introduction to the Physiology and Handling of Fruit and Vegetables*, 6th ed.; UNSW Press: Kensington, Australia; CAB International: Wallingford, UK, 2016; p. 293.
- 103. Saltveit, M.E. Effect of ethylene on quality of fresh fruits and vegetables. *Postharvest Biol. Technol.* **1999**, *15*, 279–292. [\[CrossRef\]](http://doi.org/10.1016/S0925-5214(98)00091-X)
- 104. Gwanpua, S.G.; Jabbar, A.; Tongonya, J.; Nicholson, S.; East, A.R. Measuring ethylene in postharvest biology research using the laser-based ETD-300 ethylene detector. *Plant Methods* **2018**, *14*, 105. [\[CrossRef\]](http://doi.org/10.1186/s13007-018-0372-x)
- 105. Heyes, J.A. Chilling injury in tropical crops after harvest. *Annu. Plant Rev.* **2018**, *1*, 1–31. [\[CrossRef\]](http://doi.org/10.1002/9781119312994.apr0605)
- 106. Sevillano, L.; Sanchez-Ballesta, M.T.; Romojaro, F.; Flores, F.B. Physiological, hormonal and molecular mechanisms regulating chilling injury in horticultural species. Postharvest technologies applied to reduce its impact. *J. Sci. Food Agric.* **2009**, *89*, 555–573. [\[CrossRef\]](http://doi.org/10.1002/jsfa.3468)
- 107. Beninca, T.D.T.; do Amarante, C.V.T.; Steffens, C.A.; Souza, A.G. Treatment of feijoa fruit with carnauba wax to reduce water loss and preserve postharvest quality. *Acta Hortic.* **2018**, *1205*, 919–924. [\[CrossRef\]](http://doi.org/10.17660/ActaHortic.2018.1205.118)
- 108. Biswas, P.; East, A.R.; Hewett, E.W.; Heyes, J.A. Intermittent warming in alleviating chilling injury—A potential technique with commercial constraint. *Food Bioproc. Technol.* **2016**, *9*, 1–15. [\[CrossRef\]](http://doi.org/10.1007/s11947-015-1588-7)
- 109. Al-Harthy, A.S. Postharvest Treatments to Extend the Shelf Life of Feijoa (*Acca sellowiana*). Ph.D. Thesis, Massey University, Palmerston North, New Zealand, 2010.
- 110. Amarante, C.V.T.D.; Steffens, C.A.; Benincá, T.D.T.; Hackbarth, C.; Santos, K.L.D. Quality and postharvest conservation potential of the fruit in Brazilian cultivars of feijoa. *Rev. Bras. Frutic.* **2013**, *35*, 990–999. [\[CrossRef\]](http://doi.org/10.1590/S0100-29452013000400009)
- 111. Oseko, J.; East, A.R.; Heyes, J.A. Reassessing temperature and humidity storage conditions for maintaining quality of 'Kakariki' feijoa. *Acta Hortic.* **2019**, *1256*, 157–162. [\[CrossRef\]](http://doi.org/10.17660/ActaHortic.2019.1256.22)
- 112. Zhuang, H.; Barth, M.M.; Fan, X. Respiration and browning discoloration of fresh-cut produce in modified atmosphere packaging. In *Modified Atmosphere Packaging for Fresh-Cut Fruits and Vegetables*; John Wiley & Sons, Inc.: New York, NY, USA, 2011; pp. 31–56.
- 113. Al-Harthy, A.S.; East, A.R.; Hewett, E.W.; Mawson, A.J. Controlled atmosphere storage of 'Opal Star' feijoa. *Acta Hortic.* **2010**, *876*, 401–408. [\[CrossRef\]](http://doi.org/10.17660/ActaHortic.2010.876.55)
- 114. Amarante, C.V.T.D.; Steffens, C.A.; Ducroquet, J.P.H.J.; Sasso, A. Fruit quality of feijoas in response to storage temperature and treatment with1-methylcyclopropene. *Pesqui. Agropecu. Bras.* **2008**, *43*, 1683–1689. [\[CrossRef\]](http://doi.org/10.1590/S0100-204X2008001200007)
- 115. Sganzerla, W.G.; Longo, M.; de Oliveira, J.L.; da Rosa, C.G.; de Lima Veeck, A.P.; de Aquino, R.S.; Masiero, A.V.; Bertoldi, F.C.; Barreto, P.L.M.; Nunes, M.R. Nanocomposite poly (ethylene oxide) films functionalized with silver nanoparticles synthesized with Acca sellowiana extracts. *Colloids Surf. A Physicochem. Eng. Asp.* **2020**, *602*, 125125. [\[CrossRef\]](http://doi.org/10.1016/j.colsurfa.2020.125125)
- 116. de Melo, A.P.Z.; da Rosa, C.G.; Montanheiro Noronha, C.; Machado, M.H.; Sganzerla, W.G.; da Cunha Bellinati, N.V.; Nunes, M.R.; Verruck, S.; Prudencio, E.S.; Barreto, P.L.M. Nanoencapsulation of vitamin D3 and fortification in an experimental jelly model of *Acca sellowiana*: Bioaccessibility in a simulated gastrointestinal system. *LWT* **2021**, *145*, 111287. [\[CrossRef\]](http://doi.org/10.1016/j.lwt.2021.111287)